

Curriculum development efforts for an inquiry-based introductory lab course or can we have it all?

Ayce Yesilaltay

The 7.02 Teaching Team

in alphabetical order

Instructors in Lab:

Vanessa Cheung
Ayce Yesilaltay

Instructors in Lecture:

Laurie Boyer
Mary Gehring
Piyush Gupta
Thomas Schwartz
Dane Wittrup

Writing Instructors:

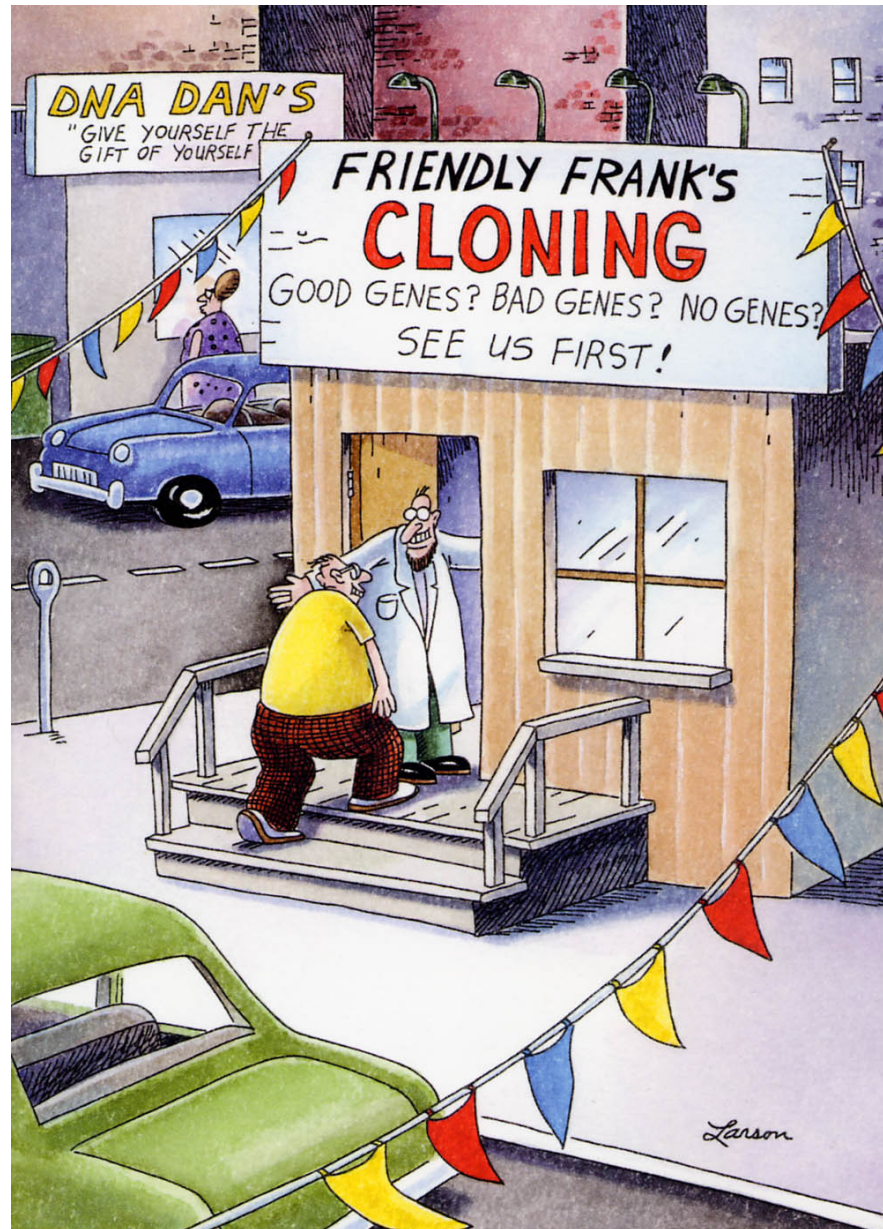
Amelia Herb
Jane Kokernak
Marilee Ogren-Balkema
Leslie Roldan

Lab Manager:

Anthony Fuccione

Graduate and undergraduate TAs

Genes and Genetics: what's not to like?



Outline

- Introduction to 7.02
- Challenges
- Development of the new curriculum
- Assessment

What is 7.02?

Introduction to Experimental Biology and Scientific Communication

- 18-unit introductory lab course
- Communications-intensive
- Prerequisite: 7.01x (Introductory Biology)
- Geared towards sophomores
- Enrollment: 110-130 students per year

Learning goals in 7.02

- Experimental techniques
 - Practice
 - Theory
 - Troubleshooting
 - Writing a lab notebook
- Data analysis
- Scientific communication
- Experimental design

Ways to learn in 7.02

all-day	22 Monday	23 Tuesday	24 Wednesday	25 Thursday	26 Friday
9 AM					
10 AM	SciComm Lecture				
11 AM		Lecture		Lecture	
Noon					
1 PM		Lab T/R	Lab W/F	Lab T/R	Lab W/F
2 PM					
3 PM					
4 PM					
5 PM					

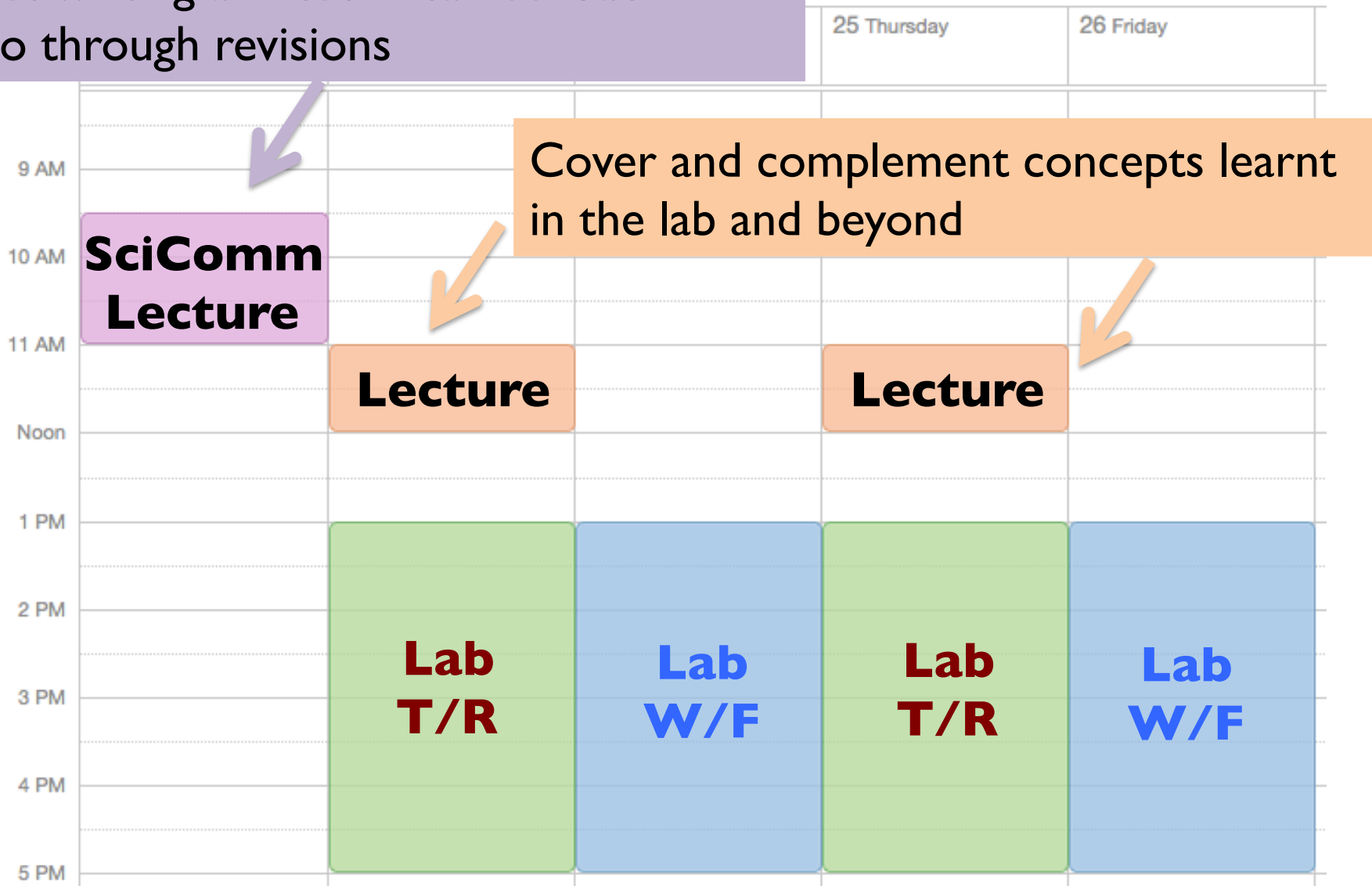
Write a manuscript on their own work
Get writing and technical feedback
Go through revisions

in 7.02

	25 Thursday		26 Friday	
9 AM				
10 AM	SciComm Lecture			
11 AM		Lecture	Lecture	
Noon				
1 PM	Lab T/R		Lab T/R	
2 PM				
3 PM	Lab W/F		Lab W/F	
4 PM				
5 PM				

Write a manuscript on their own work
Get writing and technical feedback
Go through revisions

in 7.02



Write a manuscript on their own work
Get writing and technical feedback
Go through revisions

in 7.02

25 Thursday

26 Friday

9 AM

10 AM

11 AM

Noon

Cover and complement concepts learnt
in the lab and beyond

**SciComm
Lecture**

Lecture

Lecture

Perform experiments with a lab partner
Write pre- and post lab notebook entries
Discuss in-lab questions with TA

3 PM

4 PM

5 PM

T/R

W/F

**Lab
T/R**

**Lab
W/F**

The Three Modules of 7.02

Biochemistry
(10 lab days)

Mutagenesis
Protein expression
Protein purification
Functional assays

Chemical Eng.
(4 lab days)

Yeast surface display

Genetics
(10 lab days)

Genetic screens
Recombination
Complementation
Genetic interactions

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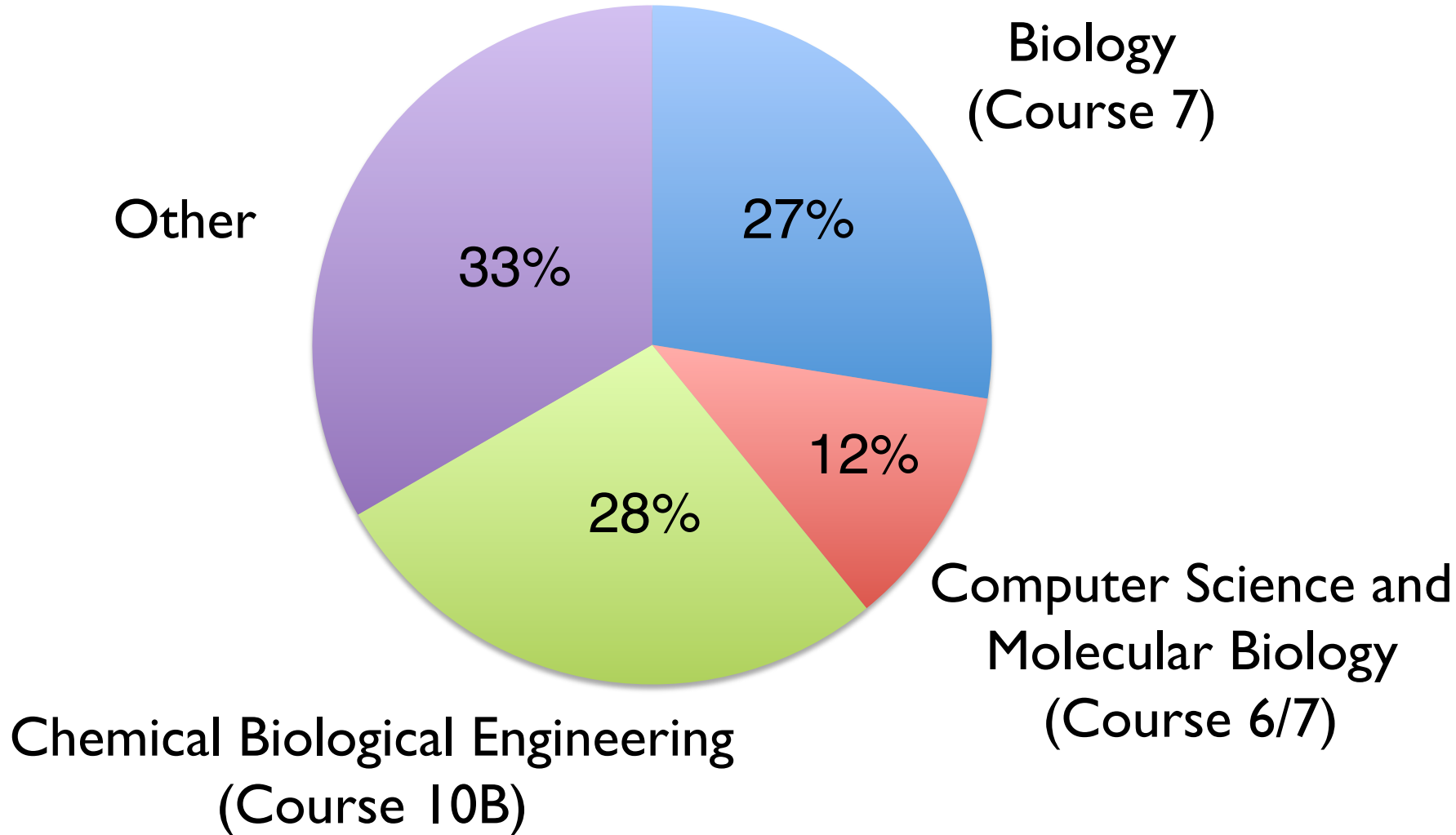
Challenges? What challenges?



Who takes 7.02?

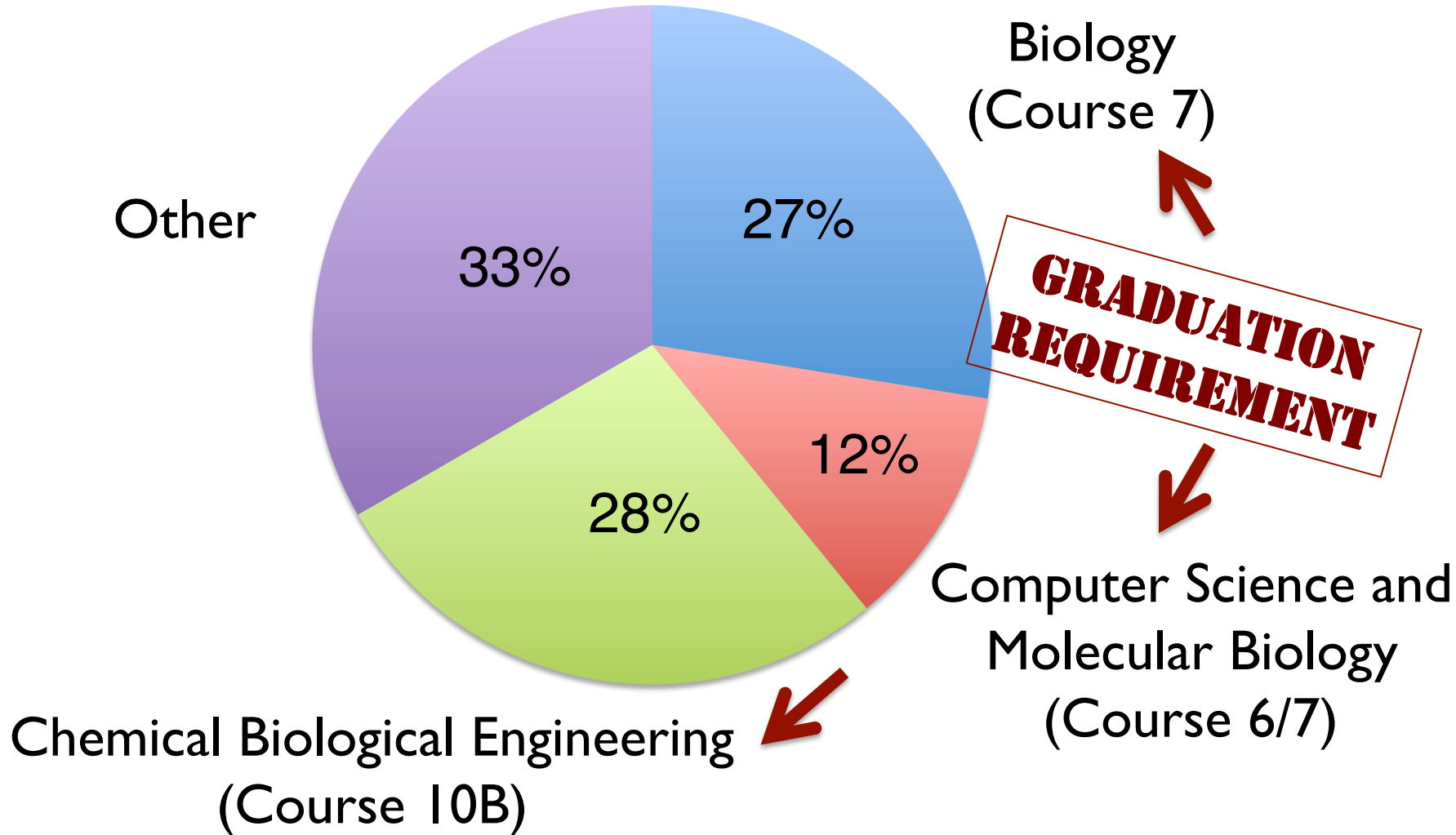


Distribution of majors in 7.02



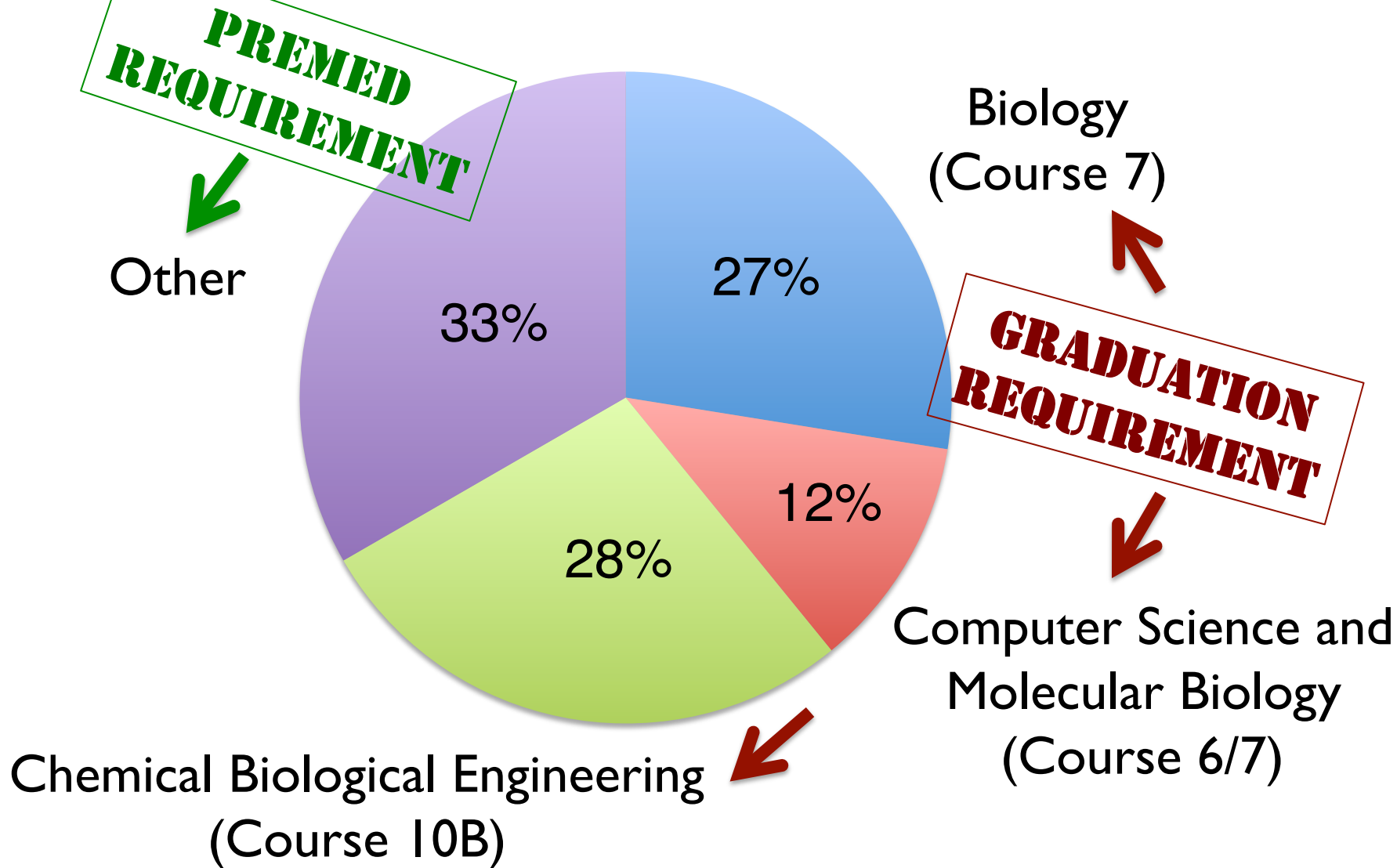
According to currently preregistered students for Fall 2014

Distribution of majors in 7.02

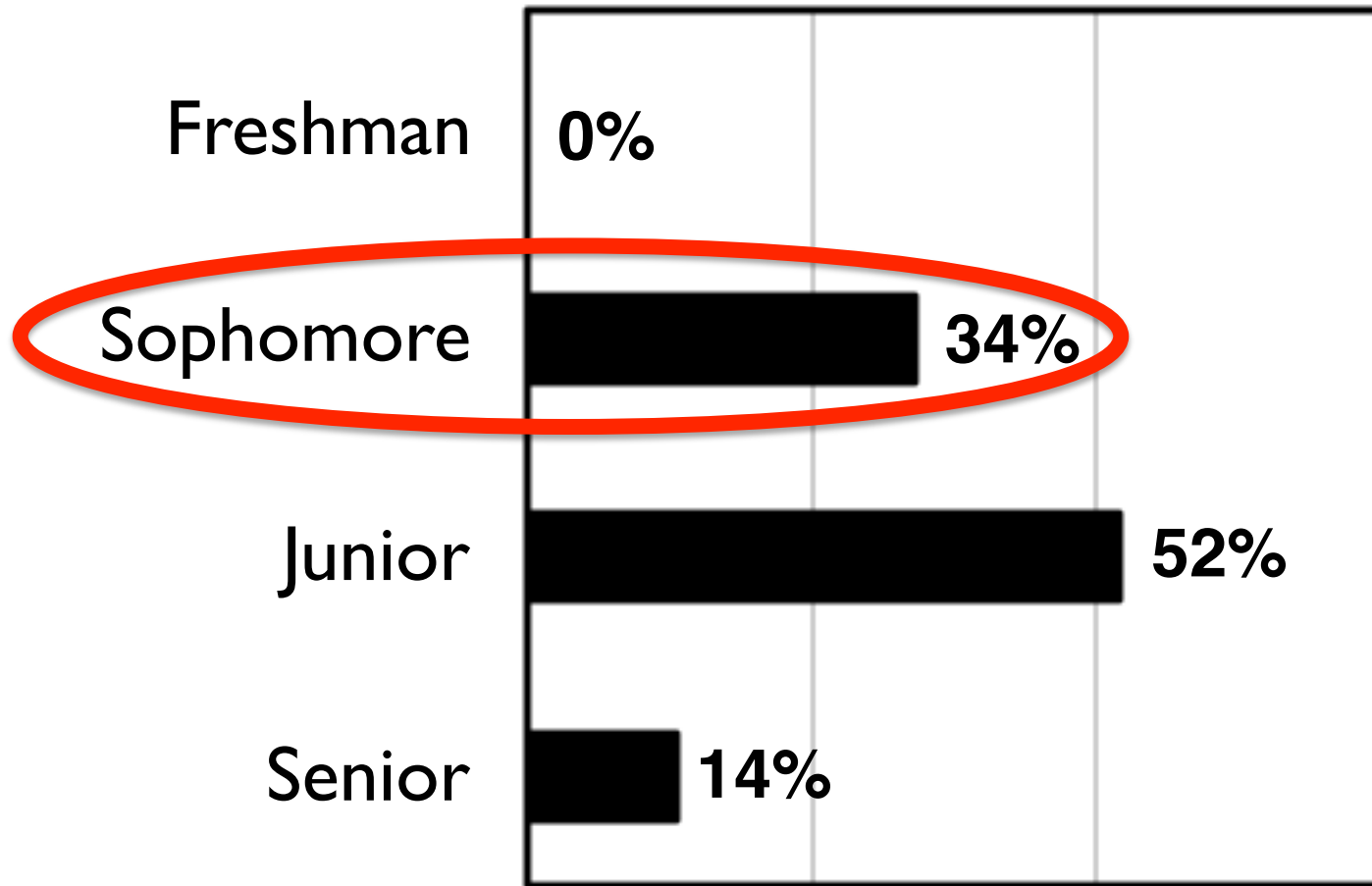


According to currently preregistered students for Fall 2014

Distribution of majors in 7.02



Which year students take 7.02?



According to currently preregistered students for Fall 2014

Challenges recapped

Preparing content for a mixed student population

- Different majors
- Different years
- Vastly varied lab experience
- Different goals and expectations

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From worm genetics to yeast genetics

What do the students think?

I hate worms!

Don't get rid of the worms! I hate yeast genetics!



Considerations for the new module

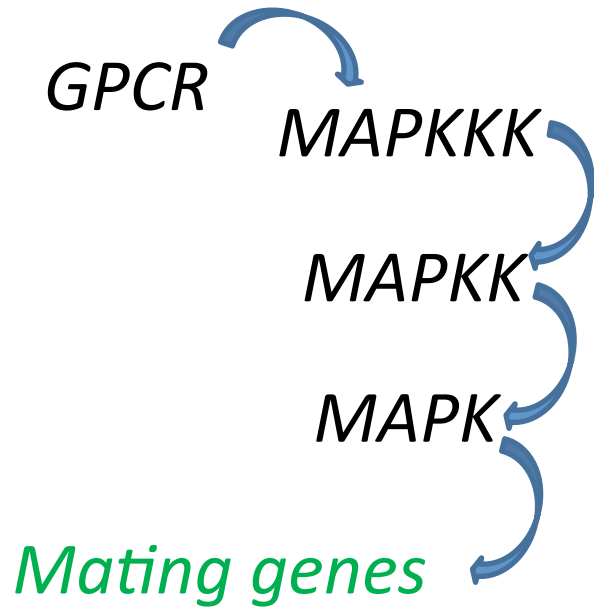
- Build the course around the genetics concepts required for the students
- Design a cohesive, hypothesis-driven module with an overarching goal
- Involve students in the decision-making process
- Potential for new research findings

The topic?

The yeast mating pathway

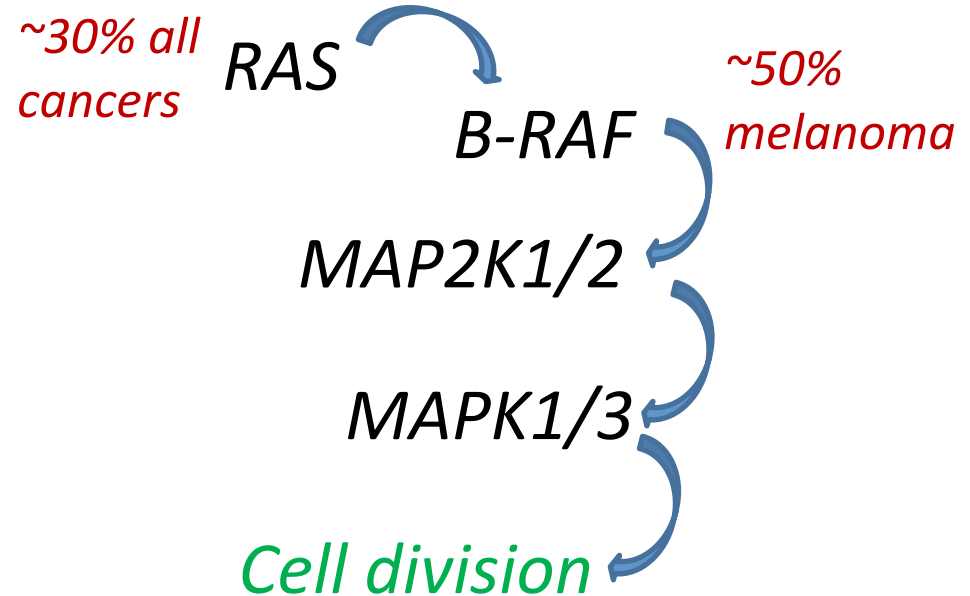
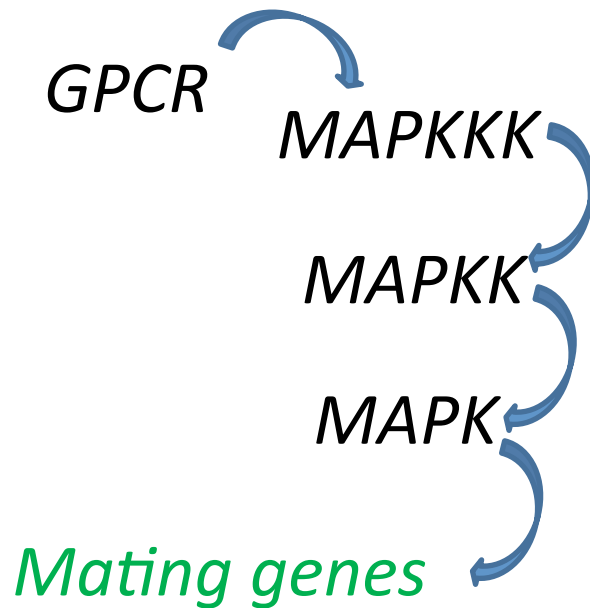
Why do we care about yeast mating?

Signal transduction is conserved



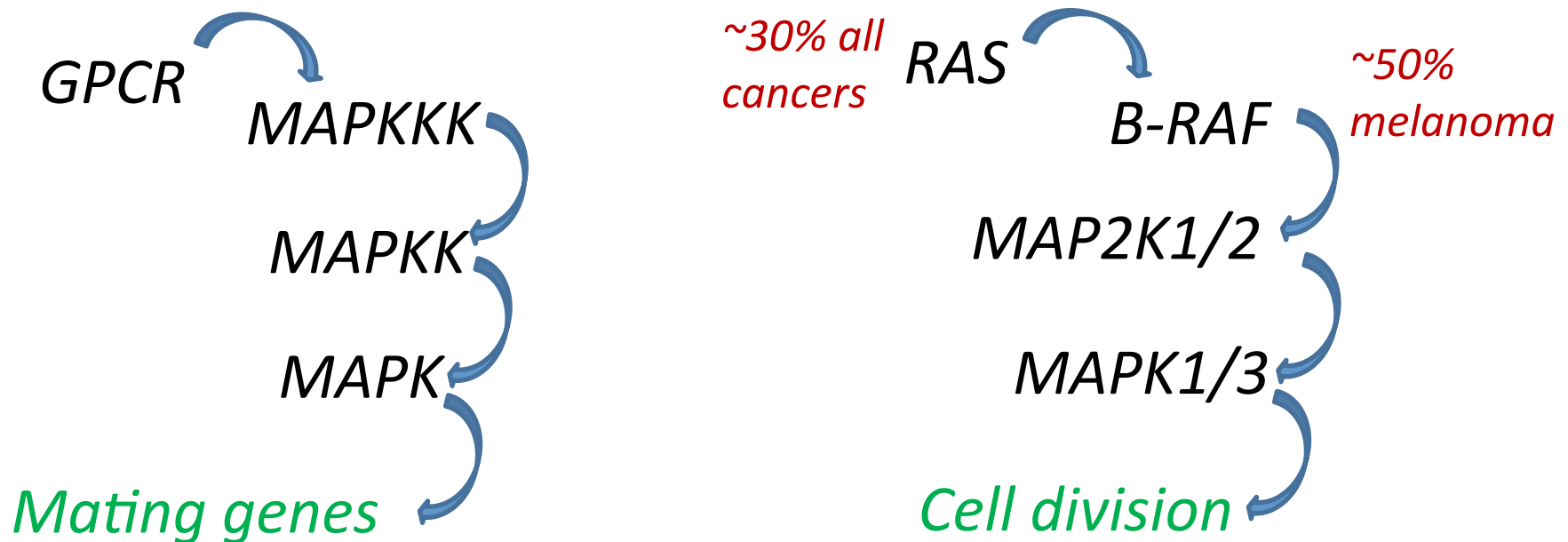
Why do we care about yeast mating?

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Why do we care about yeast mating?

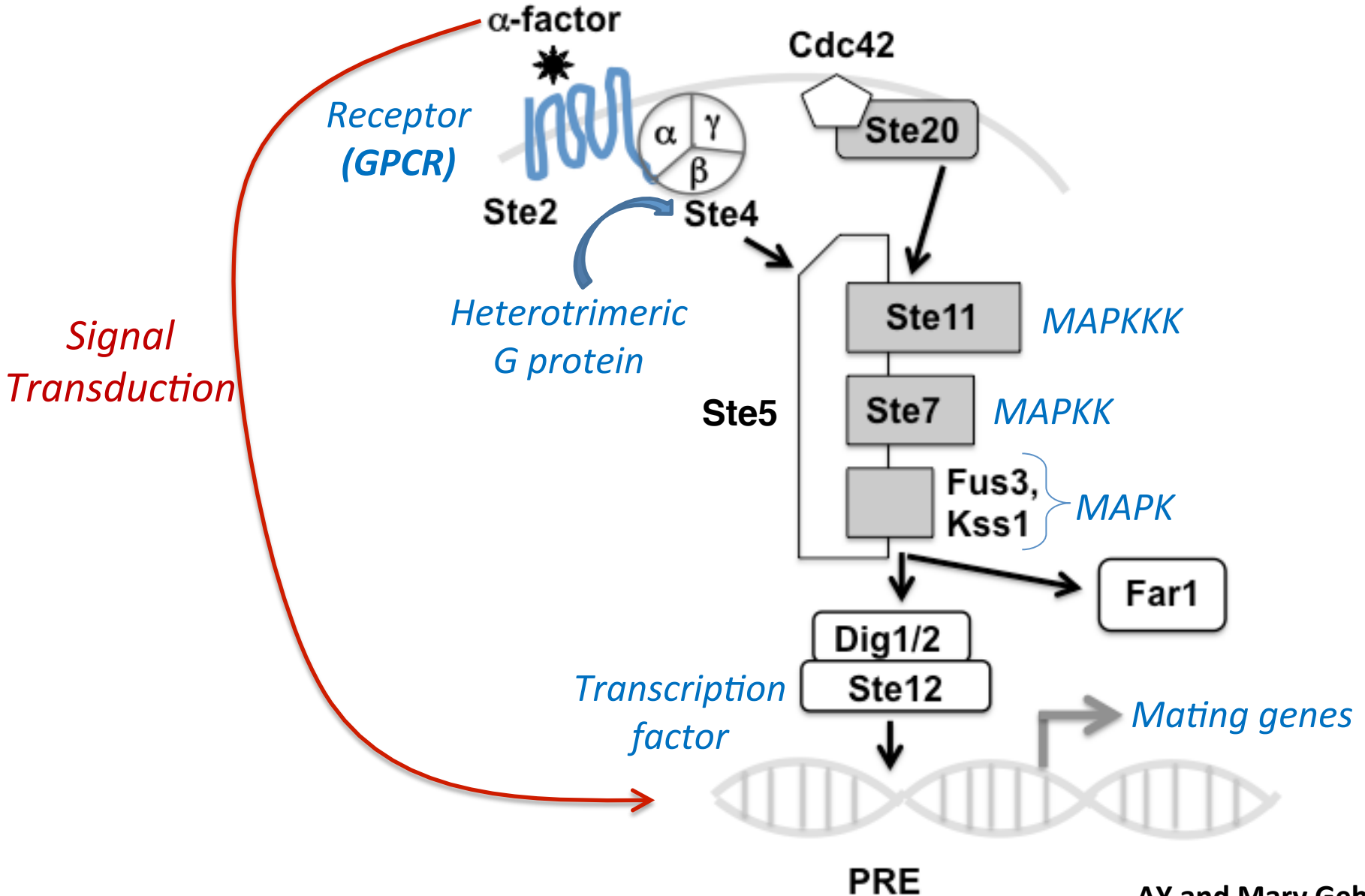
Signal transduction is conserved



Most oncogenes are mutated kinases

Many cancer drugs target kinases

Yeast mating signal transduction pathway



The new genetics module

Generate transposon-mutagenized yeast library



Select mutants defective in response to mating factor α -factor



Perform three assays to confirm the mutant phenotype



Identify the mutants by plasmid rescue, sequencing and BLAST

The new genetics module

Generation of new unique reagents



Generate transposon-mutagenized yeast library



Select mutants defective in response to mating factor α -factor



Perform three assays to confirm the mutant phenotype



Identify the mutants by plasmid rescue, sequencing and BLAST

The new genetics module

Generation of new unique reagents



Generate transposon-mutagenized yeast library

Decision making



Select mutants defective in response to mating factor α -factor



Perform three assays to confirm the mutant phenotype



Identify the mutants by plasmid rescue, sequencing and BLAST

The new genetics module

Generation of new unique reagents



Generate transposon-mutagenized yeast library



Decision making



Select mutants defective in response to mating factor α -factor



Decision making



Perform three assays to confirm the mutant phenotype



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The new genetics module

Generation of new unique reagents



Generate transposon-mutagenized yeast library



Decision making



Select mutants defective in response to mating factor α -factor



Decision making



Perform three assays to confirm the mutant phenotype



Potential for identification of new genes



Identify the mutants by plasmid rescue, sequencing and BLAST

Genes identified by the class

(and the number of times they were identified)

New gene implicated for the first time in the yeast mating pathway!

STE2

STE5 (2)

STE6

STE7 (2)

SIR3, aka STE8 (2)

SIR4, aka STE9 (3)

KCC4 (3)

TAR1 (2)

BEM4

FUS2

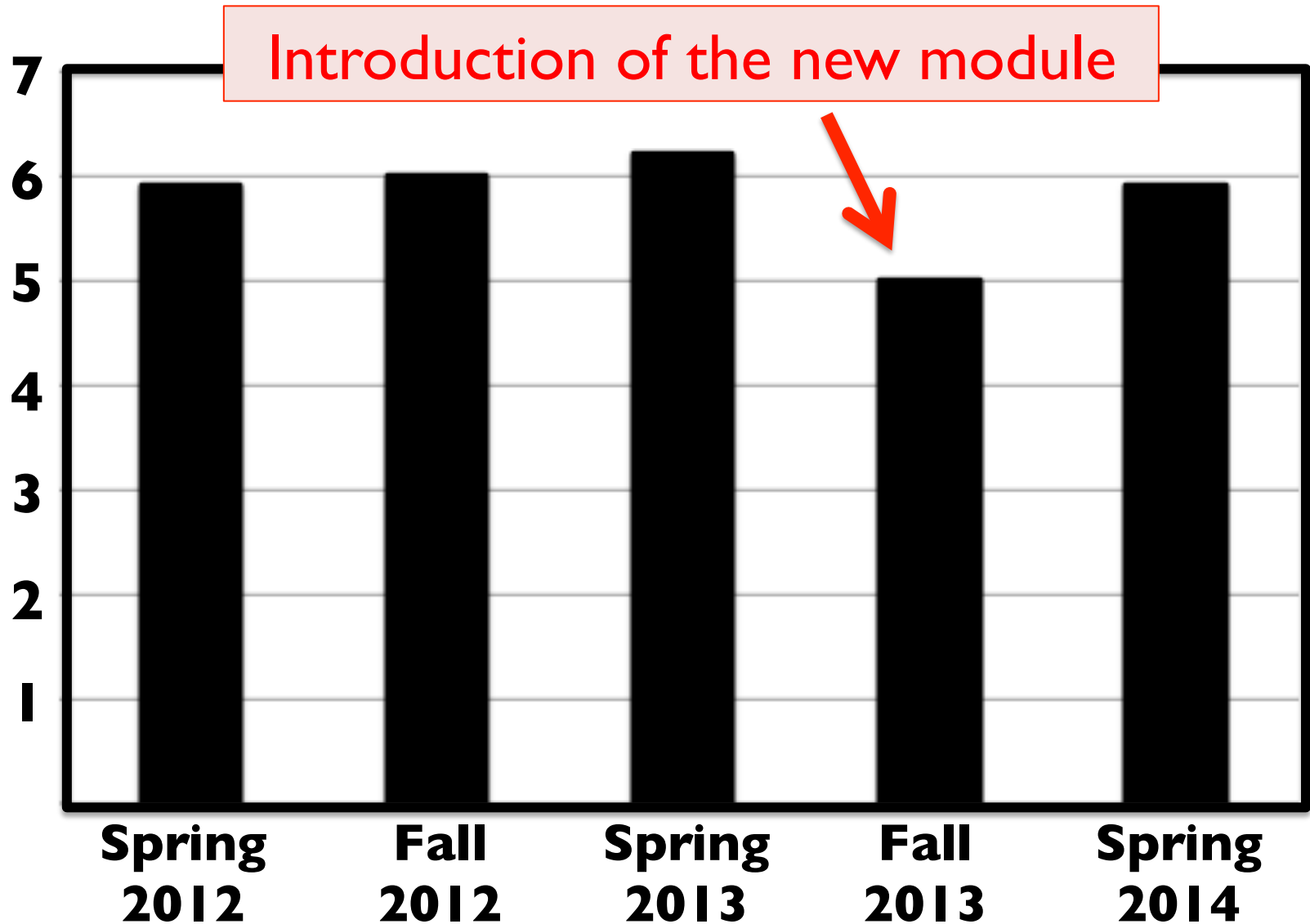
CLB6



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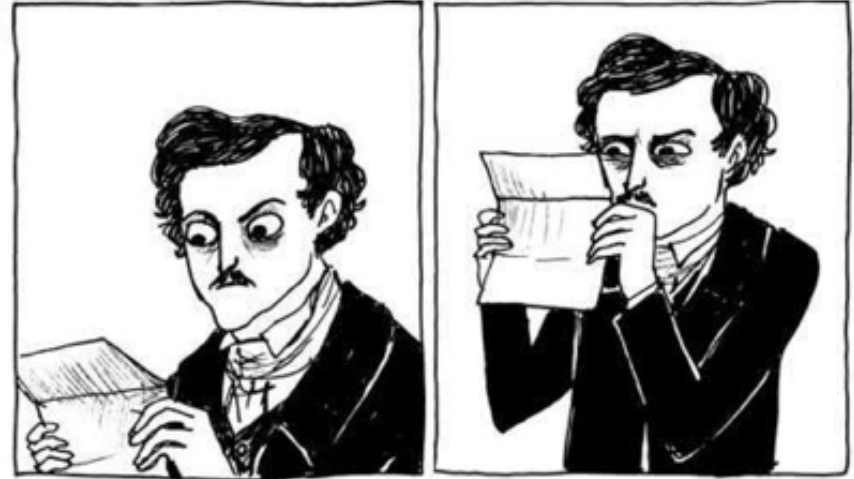
Students' subject evaluation of 7.02 (out of 7)



Paper evaluations vs online evaluations



- Higher rate of response
- More detailed feedback
- Ability to ask open-ended questions
- Ability to administer self-assessment questionnaire



What was your least favorite part of the Genetics Module?

Fall 2013

“Long lab days”

“Broken pipettes”

“Not enough spectrophotometers”

What was your least favorite part of the Genetics Module?

Fall 2013

“Long lab days”

“Broken pipettes”

“Not enough spectrophotometers”

➔ These issues were addressed for the Spring 2014 semester



Broken pipettes?

What was your least favorite part of the Genetics Module?

Spring 2014

“That one long lab day!”

Date	Day	Most students done with lab	Most students done with ILQ	Last student	Comments
2/4	Training	5:00pm			
4/8	G1				
4/9	G1	1:50pm	2:30pm	2:50pm	~30min Rec.
4/10	G 2	4:50pm	—	5:00pm	~5 left
4/11	G 2	4:50pm	—	5:05pm	~4 left
4/15	G 3	3:00pm	3:35pm	3:50pm	~4 left (30min Rec.)
4/16	G 3	3:15pm		~4:50pm	(30 min Rec.)
4/17	G 4	3:50pm	4:15pm	4:50pm	~6 left
4/18	G 4	4:15pm	4:45pm	5:00pm	
4/24	G 5			5:00pm	
4/25	G 5	4:40pm	5:00pm	5:15pm	
4/29	G 6	5:15pm		5:35pm	~8 left (~4pm start spec)
4/30	G 6	5:30pm		6:00pm	~8 left (~4:30p start spec)
5/1	G 7	~2:40pm		~3:00pm	
5/2	G 7	3:00pm	3:30pm	~3:40pm	~2 left
5/6	G 8	4:35pm		5:00pm	~5 left
5/7	G 8	4:30pm	~4:40pm	5:05pm	~4 left
5/8	G 9	2:30pm	2:50pm	3:05pm	~4 left
5/9	G 9	2:30pm	3:10pm	3:25pm	~3 left
5/13	G 10		3:00pm	~3:15pm	3:30pm Review
5/14	G 10		~3:00pm	3:30pm	3:30pm Review

What was your favorite part of the Genetics Module?

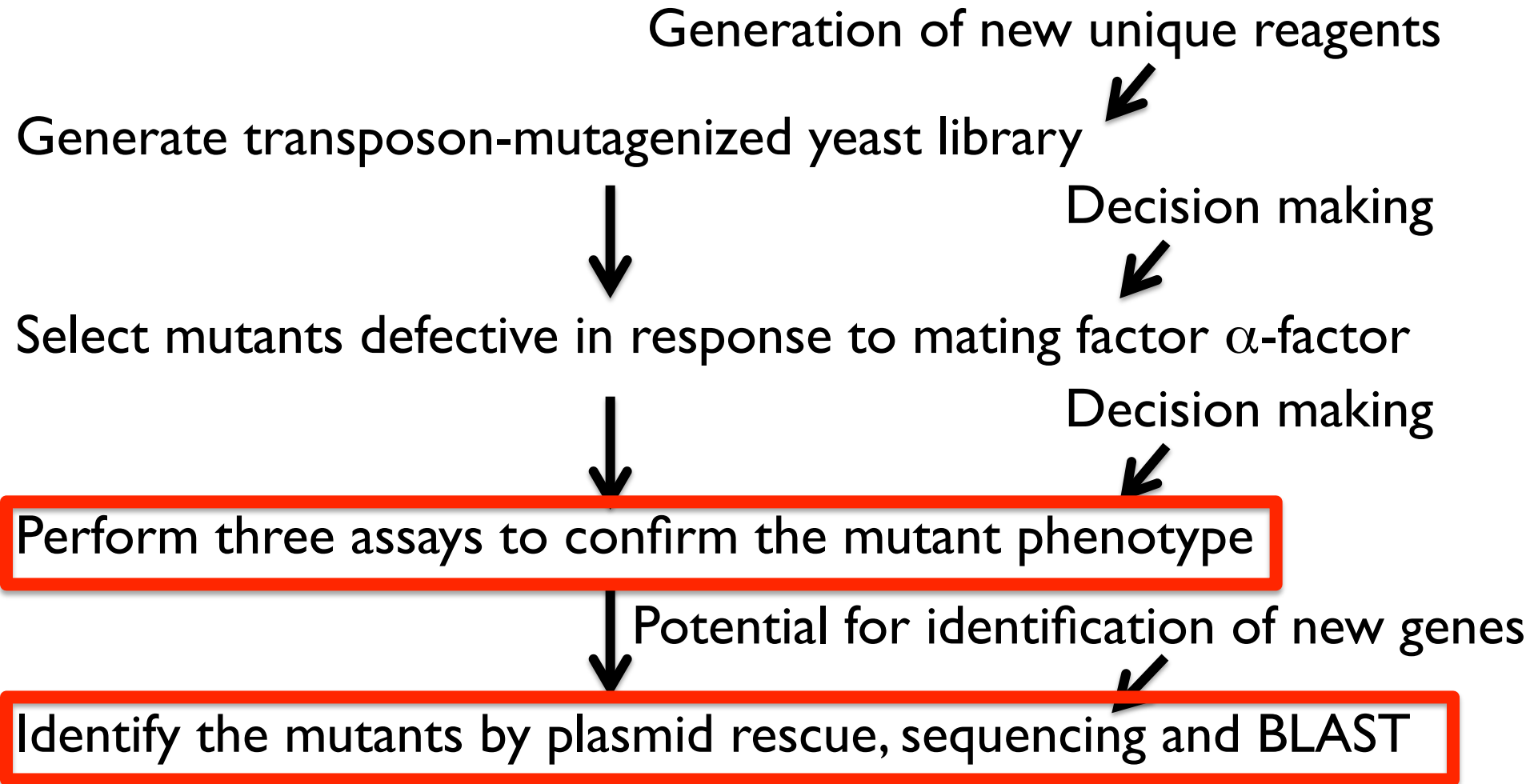
Fall 2013 and Spring 2014

“The functional assays”

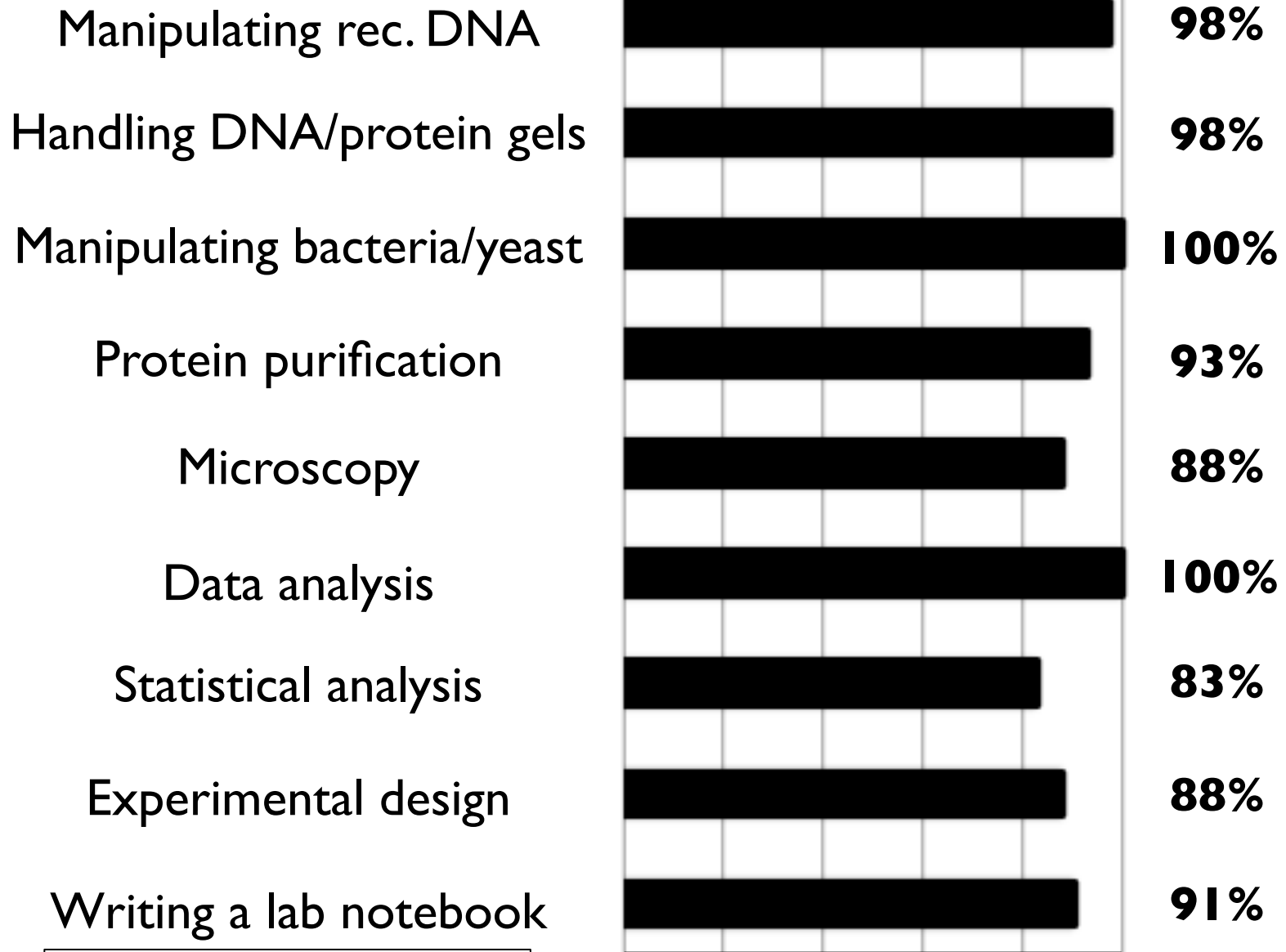
“Finding out the identity of our mutated gene”

“Seeing yeast under the microscope”

What was your favorite part of the Genetics Module?



Do you feel more comfortable with..



Response rate: 87-89%

% students who answered YES

Learning goals in 7.02

- Experimental techniques
 - Practice
 - Theory
 - Troubleshooting
 - Writing a lab notebook
- Data analysis
- Scientific communication
- Experimental design

What was the most useful course component to help your understanding?

- Practice problems
- Lectures
- Instructors
- Exams
- Review sessions
- In-lab questions
- SciComm
- Lab manual
- TAs

What was the most useful course component to help your understanding?

- TAs
- Practice problems
- Lectures
- Instructors
- Exams
- Review sessions
- In-lab questions
- SciComm
- Lab manual

TAs: our most valuable resource!

- Four graduate and eight undergraduate TAs
- Graduate TAs undergo Biology Dept. TA training
- 7.02-specific training in TA Run-Through Week
- Weekly staff meetings

Can we have it all?

- Cookbook style versus open ended
- Breadth versus depth

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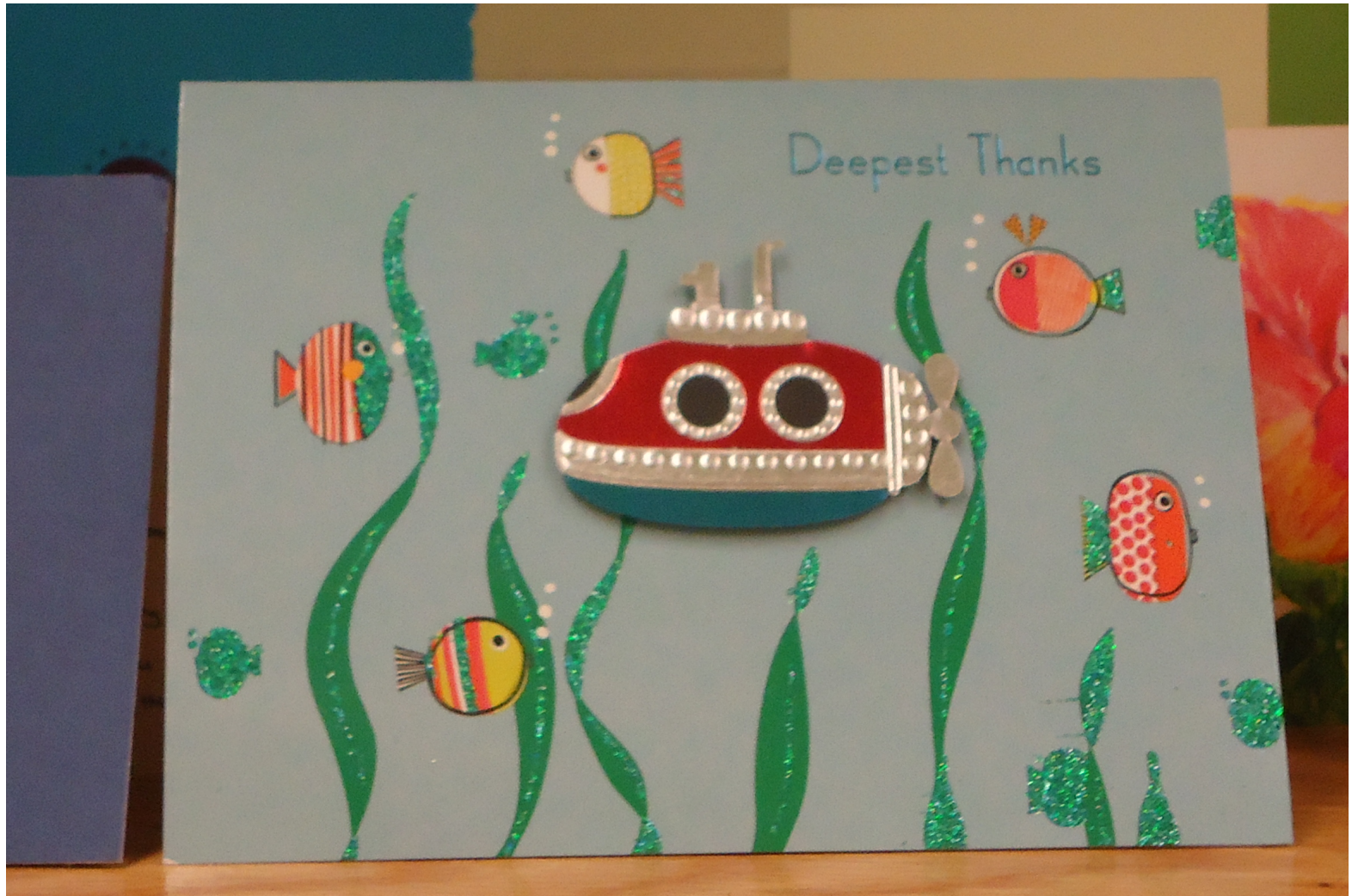
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Leslie Roldan

Lab Manager:

Anthony Fuccione

Graduate and undergraduate TAs



Thank you!

Acknowledgments

MIT

Chris Kaiser
Graham Walker
Tania Baker
Alan Grossman
Walker Lab Members
Tom RajBhandary
Caroline Kohrer
Mary Ellen Wiltrout
Mandana Sassanfar
Beth Vogel-Taylor
Steve Bell
Monty Krieger
Krieger Lab Members
Alison Brauneis
Lourdes Aleman
Bob Horvitz
Wendy Gilbert
Lenny Guarante
Glenda Stump
Courtney Crummett
Wendy Salmon

UMass Worcester

Peter Pryciak
Duane Jenness

Stanford University

Mike Snyder
Tim Stearns

University of Michigan

Anuj Kumar

SUNY

Jamie Konopka
Deborah Spikes

Suffolk University

Melanie Berkmen

Princeton University

Allison Gammie

Brandeis University

Melissa Kosinski-Collins

Boston University

Meredith Knight

Funding

MIT DEPARTMENT OF
BIOLOGY

HHMI
HOWARD HUGHES MEDICAL INSTITUTE

The Alumni Class Funds
supporting education at MIT since 1994

Stay tuned for the next curriculum...

